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Carbohydrate-protein interactions are crucial for a multitude of biological functions. Beginning from the discovery of the multivalent effects as a central theme few decades ago, the roles of carbohydrate-protein interactions are constantly uncovered in the larger context of biological importance. Whereas many finer details of the interactions are delineated well to an isolated protein and ligand, circumstances at which these interactions occur in cell membrane systems are not known in detail currently in literature at large. An objective of the present study is to identify the factors governing the nature of interactions with the aid of well-defined synthetic structures that mimic the cellular environments. Vesicles that represent the cell surfaces are utilized in the present study to uncover the finer mechanisms of the macromolecular ligand-receptor interactions. Sugar ligands are incorporated in varying densities at the surfaces of synthetic vesicles, in order to investigate their protein binding patterns. A carbohydrate binding protein, namely, concanavalin A, is studied for its interaction with cognate high affinity \Box -D-mannopyranoside ligand present at the vesicle surfaces. The nature of interaction follows strictly the ligand-densities at the vesicle surface. A major outcome of the investigations is that the vesicles having sparsely populated ligands engage in an interaction, leading to cross-linked, multimeric ligand-lectin complex formation. Whereas the vesicles having mostly or fully populated ligands promote only an interaction which is monomeric in nature, namely, one lectin tetramer interacting with one glycovesicle on the same plane of the vesicle. Multimeric aggregate formation does not accrue when the vesicle is mostly or fully populated with the sugar ligands. The study is put investigations through varied techniques, relating to changes in the hydrodynamic diameters, particle sizes, secondary structures and the emission behavior of ligand-receptor complexes. The efficiency of binding occurs in nanomolar dissociation constants, within which the monomeric glycovesicle-lectin interaction possesses relatively tighter binding than the multimeric complex. The ability to fine-tune the modes of ligand-receptor interactions has a fundamental significance to advancing the therapeutic regimes, particularly in the circumvention of disease progressions originating from intra- and intercellular communication processes. The observations of the present study that ligand densities are the sources to alter the modes of receptor binding bear a greater significance to advance the frontiers in ligand-receptor interactions in general and in carbohydrate mediated biological processes in particular.